

# Article



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# Enhanced walking-induced fat oxidation by New Zealand blackcurrant extract is body composition-dependent in recreationally active adult females

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Abstract: New Zealand blackcurrant (NZBC) extract enhanced cycling-induced fat oxidation in fe-11 male endurance athletes. We examined in recreationally active females the effects of NZBC extract 12 on physiological and metabolic responses by moderate-intensity walking and the relationship of fat 13 oxidation changes with focus on body composition parameters. Twelve females (age: 21±2 yrs, BMI: 14 23.6±3.1 kg·m<sup>-2</sup>) volunteered. Bioelectrical bioimpedance analysis was used for body composition 15 measurements. Resting metabolic equivalent (1-MET) was 3.31±0.66 ml·kg<sup>-1</sup>·min<sup>-1</sup>. Participants com-16 pleted an incremental walking test with oxygen uptake measurements to individualize the treadmill 17 walking speed at 5-MET. In a randomized, double-blind, cross-over design, the 30-min morning 18 walks were in the same phase of each participant's menstrual cycle. No changes by NZBC extract 19 were observed for walking-induced heart rate, minute ventilation, oxygen uptake and carbon diox-20 ide production. NZBC extract enhanced fat oxidation (10 responders, range: 10-66%). There was a 21 significant correlation for changes in fat oxidation with body mass index, body fat% in legs, arms 22 and trunk, and a trend with fat oxidation at rest but not with body mass and habitual anthocyanin 23 intake. The NZBC extract responsiveness of walking-induced fat oxidation is body composition-24 dependent and higher in young-adult females with higher body fat% in legs, arms and trunk. 25

Keywords: anthocyanins; substrate oxidation; exercise; body composition

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# 1. Introduction

The energy requirements of moderate-intensity exercise are provided primarily by 29 the oxidation of carbohydrates and lipids [1]. The relative contribution by the oxidation 30 of carbohydrates and lipids towards the moderate-intensity exercise requirements are af-31 fected by dietary intake [2], training status [3], sex [4], exercise modality [5], environmen-32 tal conditions [6], and supplementation (e.g. caffeine [7], green tea extract [8], Matcha 33 green tea [9]; New Zealand blackcurrant [10]). Many studies have examined the factors 34 that contribute to exercise-induced maximal fat oxidation. For example, unfit women with 35 obesity have a higher maximal fat oxidation than unfit women with normal weight [11]. 36 However, observations that body composition may affect exercise-induced fat oxidation 37 are not consistent. Kerhervé et al [12] observed that body composition differences be-38 tween women did not affect exercise-induced fat oxidation but noted substantial inter-39 individual differences in normal weight, overweight and obese women. In addition, fat 40 mass localization may affect exercise-induced fat oxidation [13]. However, studies on the 41 effect of supplementation-induced fat oxidation have not addressed the potential effect of 42 regional fat distribution. Women with similar body mass and body mass index may still 43

vary substantially in the localisation of body fat. In general, enhanced fat oxidation by 44 supplementation is considered beneficial with implications for individuals with weight 45 management issues. For example, Moro orange extracts seems to have lipolytic effects and 46 reduces abdominal fat [14]. Although a substantial reduction of energy intake is most ef-47 fective for weight loss (e.g. [15]), dietary choices may be helpful for weight management. 48 In addition, for the general population, there is an interest to affect the exercise-induced 49 metabolic responses to get enhanced health benefits. Many supplements can stimulate li-50 polysis [16]. Enhanced fat oxidation by supplement use can be partly due to enhanced 51 lipolysis with the overall metabolic effect decided by the summed effects of regional lipol-52 ysis. In Şahin et al [17], effects of New Zealand blackcurrant extract on whole-body fat 53 oxidation by moderate-intensity walking was affected by body composition in recreation-54 ally active males. In males, higher body mass index and body fat% was providing higher 55 levels of enhanced whole-body exercise-induced fat oxidation [17]. Whether such obser-56 vations can be transferred to women is not known. In addition, in Şahin et al [17], the 57 relationship for regional body fat% with enhanced fat oxidation was not examined. There-58 fore, the main aim of the present is to examine the effects of intake of New Zealand black-59 currant extract on physiological and metabolic responses by moderate-intensity treadmill 60 walking in recreationally active women. In addition, we will primarily examine whether 61 there is a relationship between the blackcurrant-induced changes in whole-body fat oxi-62 dation and body fat% in upper limb, lower limbs, and trunk. 63

# 2. Materials and Methods

## 2.1. Participants

The female participants (n=12, age: 21±2 yr; height: 166±7 cm; body mass: 65±11 kg; 66 BMI: 23.7±3.1 kg·m<sup>-2</sup>) recruited for the study were healthy recreationally active Caucasian 67 University students and staff. All females had a regular menstrual period, were non-68 smokers, and were not taking other dietary supplements. Participants provided written 69 informed consent and health status was confirmed by a health history questionnaire. The 70 physical activity level of the female participant's physical activity level was quantified 71 with the short version of the International Physical Activity Questionnaire (2749±2106 72 MET· week-1) [18]. The present study accepted the following methods of contraception: 73 combined pill, diaphragm or intrauterine device. Approval for the study was obtained 74 from the University of Chichester Research Ethics Committee (ethical approval code: 75 1819\_1600944). 76

# 2.2. Experimental design

The study had a randomized, placebo-controlled, cross-over design. Female partici-79 pants visited the laboratory for one preliminary and two experimental sessions. For each 80 session, participants abstained from unaccustomed and intense exercise for 48 h, had no 81 alcohol and caffeine intake for 24 h, and were at least 2-hour postprandial after consuming 82 a breakfast of one slice of bread and a glass of water [10]. Sessions were in the morning 83 with a month between the two experimental sessions to ensure that the females were 84 tested in the same phase of the participant's menstrual cycle [19]. The participants were 85 supplemented with placebo or NZBC extract (see below for details) for seven days before 86 each experimental session [10, 20]. Participants completed a food frequency questionnaire 87 with anthocyanin-containing foods and drinks listed in the Phenol-Explorer database [21], 88 to estimate daily habitual anthocyanin intake (32±26 mg·day-1; range: 5 to 88 mg·day-1).

## 2.3. Preliminary session

In the preliminary session, height (Seca 213, Seca, Birmingham, United Kingdom), 92 and body mass (Kern ITB, Kern, Germany) were measured as well as body composition 93 (Tanita BC418, segmental body composition analyzer, Tanita, Illinois, USA). Subsequently, participants were fitted with a heart rate monitor (Polar F1, Polar Electro (UK) 95 Ltd, Warwick, United Kingdom). Participants then rested for 30 minutes in a chair, 96

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followed by 2x10 minutes expired air collection using Douglas bags (Cranlea & Co. 97 Bourneville, Birmingham, UK) to determine the oxygen consumption at rest. The lowest 98 value for oxygen consumption at rest was taken as the one metabolic equivalent (1-MET) 99 (1-MET: 3.31±0.66 ml·kg<sup>-1</sup>·min<sup>-1</sup>). Heart rate at rest was 69±9 beats min<sup>-1</sup>. Blood pressure at 100 rest was taken twice (OMRON 705 IT, Medisave, Weymouth, UK) and averaged (systolic 101 blood pressure: 120±8 mmHg, diastolic blood pressure: 70±7 mmHg). Subsequently, par-102 ticipants completed an incremental-intensity walking protocol on a treadmill (Woodway 103 Ergo ELG 70, Cranlea & Co. Bourneville, Birmingham, UK). Treadmill incline was 1%. 104 Participants completed 5x8-minute stages starting at 2 km·h<sup>-1</sup> progressing by 1 km·h<sup>-1</sup> until 105 a speed of 6 km·h-1 was reached [22]. In the last 3-min of each 8-min stage, expired air was 106 collected using Douglas bags. Expired air was analyzed for fractions of oxygen and carbon 107 dioxide by a 3-point calibrated gas analyser (Series 1400, Servomex, Crowborough, East 108Sussex, UK), and volume was measured (Harvard Apparatus Ltd., Edenbridge, UK). Ex-109 pired gas volumes were corrected to standard temperature and pressure and dry gas con-110 ditions and calculated using Haldane transformation with consideration of inspired frac-111 tions of oxygen and carbon dioxide that were measured each time halfway during each 3-112 min expired air collection. The incremental-intensity walking protocol with measurement 113 of oxygen consumption was performed to establish for each participant the linear rela-114 tionship between walking speed and the metabolic equivalent ( $r^{2}=0.9626\pm0.0259$ ). The lin-115 ear relationship between walking speed and metabolic equivalent allowed for each par-116 ticipant to establish the walking speed at 5-METs (i.e. moderate-intensity exercise, walk-117 ing speed: 5.53±0.39 km·h<sup>-1</sup>). 118

#### 2.4. Supplementation for the experimental sessions

For the experimental sessions, two capsules of NZBC extract (one capsule contain-121 ing 300 mg active cassis, of which 105 mg were anthocyanins, i.e., 35–50% delphinidin-3-122 O-rutinoside, 5–20% delphinidin-3-O-glucoside, 30–45% cyanidin-3-O-rutinoside, 3–10% 123 cyanidin-3-O-glucoside) (CurraNZ, Health Currancy Ltd., Surrey, United Kingdom) or 124 placebo pills (2×300 mg microcrystalline cellulose M102) were taken for 7 days. The final 125 two capsules were taken 2 hours before visiting the laboratory for the experimental ses-126 sions. Participants were allowed a breakfast consisting of one slice of bread and a glass of 127 water 3 hours before visiting for the experimental sessions. 128

#### 2.5. Experimental sessions

For the experimental sessions, participants completed a 30 min moderate-intensity 131 treadmill walk (4.7±0.4 METs) at the speed established in the preliminary session with 132 recording of heart rate (Polar F1, Polar Electro (UK) Ltd, Warwick, United Kingdom) and 133 expired air collection from 7-10, 17-20 and 27-30 min. Rates of whole body fat and carbo-134 hydrate oxidation were calculated with equations below from Jeukendrup and Wallis [23] 135 and the assumption of negligible protein oxidation: 136

Fat oxidation (g × min<sup>-1</sup>) = 
$$1.695 \times \dot{V}O_2 - 1.701 \times \dot{V}CO_2$$

Carbohydrate oxidation (g × min<sup>-1</sup>) =  $4.210 \times \dot{V}CO_2 - 2.962 \times \dot{V}O_2$ 

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The respiratory exchange ratio was calculated by dividing the volume of carbon dioxide produced by the volume of oxygen consumed.

#### 2.6. Statistical analysis

Statistical analyses were completed using Graphpad Prism 5 for Windows. Physiological and metabolic responses by the moderate-intensity walk were measured from 7-10, 17-20 and 27-30 min during the walk and averaged. All parameters were tested for normality with the D'Agostino and Pearson omnibus test. Two-tailed paired sample t-150

tests were used to compare all the parameters between the placebo and NZBC extract 151 conditions. Values are reported as mean±SD and 95% confidence intervals. Statistical sig-152 nificance was accepted at P < 0.05. P-values of  $0.05 \ge P \le 0.1$  were interpreted according to 153 guidelines by Curran-Everett and Benos [24]. Cohen's d effect sizes were calculated and 154 considered trivial (d < 0.2), small (d = 0.2-0.49), moderate (d = 0.5-0.79) and large (d  $\ge 0.8$ ). 155 Pearson correlation coefficients were calculated and tested for significance for the rela-156 tionships between habitual anthocyanin intake (not considering the NZBC extract intake), 157 fat oxidation at rest, body mass, body mass index, fat % of the legs, arms and trunk and 158 changes of walking-induced fat oxidation with intake of NZBC extract in comparison to 159 walking-induced fat oxidation in the placebo condition. The  $\Delta$  FAO is the fat oxidation 160 with intake of New Zealand blackcurrant extract minus the fat oxidation with intake of 161 placebo. The required sample size was not calculated and the number of 12 participants 162 was lower than previous studies with observations of an effect of NZBC extract on exer-163 cise-induced fat oxidation [males, n=14 [10], males, n=15 [17, 25]), females, n=16 [26]). 164

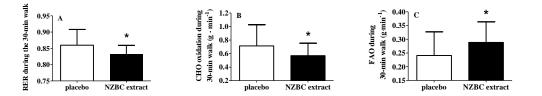
# 3. Results

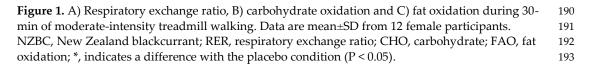
# 3.1. Walking-induced physiological responses

During the moderate-intensity treadmill walk, the NZBC extract had no effect on 167 heart rate (PL: 121±17 beats·min<sup>-1</sup>, 95% CI [110, 132 beats·min<sup>-1</sup>]; NZBC extract: 120±14 168 beats·min<sup>-1</sup>, 95% CI [111, 129 beats·min<sup>-1</sup>]; P=0.58, d= -0.06), minute ventilation (PL: 25.8±7.4 169 L·min<sup>-1</sup>, 95% CI [21.1, 30.5 L·min<sup>-1</sup>]; NZBC extract: 24.4±6.3 L·min<sup>-1</sup>, 95% CI [20.5, 28.4 170 L·min<sup>-1</sup>], P=0.14, d= -0.20), oxygen uptake (PL: 16.1±2.1 mL·kg<sup>-1</sup>·min<sup>-1</sup>, 95% CI [14.8, 17.5 171 mL·kg<sup>-1</sup>·min<sup>-1</sup>; NZBC extract: 15.8±1.8 mL·kg<sup>-1</sup>·min<sup>-1</sup>, 95% CI [14.6, 16.9 mL·kg<sup>-1</sup>·min<sup>-1</sup>], 172 P=0.45, d= -0.18) and carbon dioxide production (PL: 13.8±2.1 mL·kg<sup>-1</sup>·min<sup>-1</sup>, 95% CI [12.5, 173 15.1 mL·kg<sup>-1</sup>·min<sup>-1</sup>]; NZBC extract: 13.1±1.4 mL·kg<sup>-1</sup>·min<sup>-1</sup>, 95% CI [12.2, 14.0 mL·kg<sup>-1</sup>·min<sup>-</sup> 1741], P=0.17, d= -0.40). The absence of an effect on the physiological responses by NZBC ex-175 tract indicate no change in the regulatory mechanism for exercise-induced heart rate and 176 respiratory demands. 177

#### 3.2. Walking-induced metabolic responses

New Zealand blackcurrant extract provided 3.0% lower values for RER (PL: 95% CI 179 [0.83, 0.89], NZBC extract: 95% CI [0.82, 0.85], P=0.009, d= -0.69) (Figure 1A), 10.8% lower 180 values for carbohydrate oxidation (PL: 95% CI [0.51, 0.91 g·min<sup>-1</sup>], NZBC: 95% CI [0.45, 181  $0.69 \text{ g·min}^{-1}$ , P=0.03, d= -0.56) (Figure 1B) and 25.0% higher values for fat oxidation (PL: 182 95% CI [0.19, 0.30 g·min<sup>-1</sup>], NZBC extract: 95% CI [0.24, 0.34 g·min<sup>-1</sup>], P=0.005, d=0.59) (Fig-183 ure 1C). The 10 participants (~83%) with increased walking-induced fat oxidation re-184 sponded by an average of 32% (SD: 17%, range: 10-66%) with 9 participants higher than 185 14%. The changes in metabolic responses by moderate-intensity walking with intake of 186 NZBC extract indicate a change in the regulation of exercise-induced carbohydrate and 187 fat metabolism. 188





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#### 3.3. Habitual anthcoyanin intake, fat oxidation at rest and walking-induced fat oxidation

There was no significant correlation between habitual dietary anthocyanin intake 195 and the absolute change in walking-induced whole-body fat oxidation (r<sup>2</sup>=0.04, P=0.56) 196 (Figure 2A). The habitual dietary intake of anthocyanins did not include the anthocya-197 nins consumed by intake of the NZBC extract. The absence of a relationship between 198 habitual dietary intake and the absolute change in exercise-induced fat oxidation sug-199 gest that the enhanced metabolic response by intake of NZBC extract was not due to a 200 low habitual anthocyanin intake. There was a trend for a significant correlation between 201 fat oxidation at rest and the absolute change in walking-induced fat oxidation (r<sup>2</sup>=0.26, 202 P=0.09) (Figure 2B). This suggest an enhanced response for exercise-induced fat oxida-203 tion with intake of NZBC extract for females with higher whole-body fat oxidation at 204 rest. 205

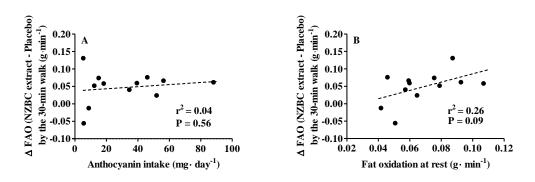


Figure 2. Relationship between habitual anthocyanin intake (A) and fat oxidation at rest (B) and208the changes in fat oxidation ( $\Delta$  FAO) by 30-min of moderate-intensity treadmill walking with in-209take of New Zealand blackcurrant (NZBC) extract. The  $\Delta$  FAO is the fat oxidation with intake of210New Zealand blackcurrant extract minus the fat oxidation with intake of placebo. The habitual211anthocyanin intake did not include the anthocyanin intake by New Zealand blackcurrant extract.212

#### 3.4. Body mass, body composition and walking-induced fat oxidation

There was no significant correlation between body mass and the absolute change in 214 walking-induced fat oxidation ( $r^2=0.23$ , P=0.11) (Figure 3A). However, there was a significant correlation between body mass index ( $r^2=0.53$ , P=0.008) (Figure 3B), body fat% of the 216 legs ( $r^2=0.57$ , P=0.005) (Figure 3C), body fat% of the arms ( $r^2=0.46$ , P=0.016) (Figure 3D) and 217 body fat% of the trunk ( $r^2=0.44$ , P=0.019) (Figure 3E) and the absolute change in whole-218 body fat oxidation. 219

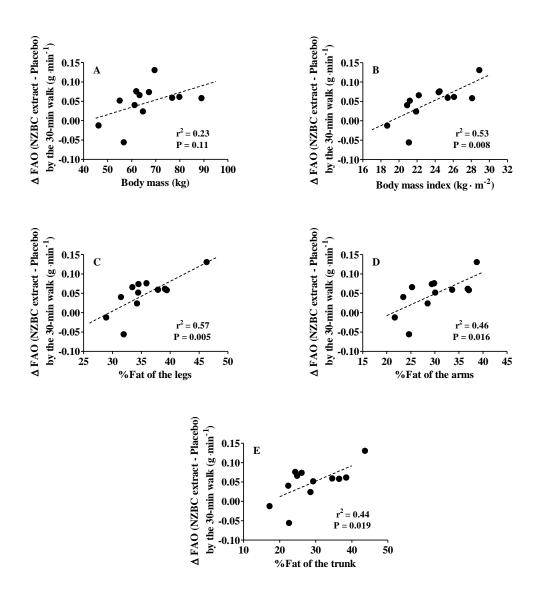
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Figure 3. Relationship between body mass (A), body mass index (B), %fat of the legs (C), %fat of224the arms (D) and %fat of the trunk (E) and the changes in fat oxidation ( $\Delta$  FAO) by 30-min of mod-225erate-intensity treadmill walking with intake of New Zealand blackcurrant (NZBC) extract. The  $\Delta$ 226FAO is the fat oxidation with intake of New Zealand blackcurrant extract minus the fat oxidation227with intake of placebo.228

The observations on body composition parameters suggest an enhanced response for 229 exercise-induced fat oxidation with intake of NZBC extract for females with higher body 230 fat in the legs, arms and trunk. The significant correlations for the relationship between 231 body mass index, body fat % in the legs, arms and trunk and changes in walking-induced 232 fat oxidation may suggest differences in adipocyte sensitivity in response to intake of 233 NZBC extract in females with higher body fat%. 234

# 4. Discussion

The present study presents novel findings on the effect of intake of anthocyanin-rich236New Zealand blackcurrant extract on the physiological and metabolic responses during237moderate-intensity walking in recreationally active females. Recently, Elliott-Sale et al238[27] emphasized the need for inclusion of women as participants in exercise science239studies. Our findings contribute to the limited information that is available on the240

ergogenic effects during exercise by intake of a berry supplement in studies with only 241 female participants. Previous studies have reported on the enhanced exercise-induced 242 fat oxidation by intake of New Zealand blackcurrant extract (7 days, 210 mg anthocya-243 nins per day) in recreationally active males during moderate-intensity walking [20] and 244 endurance-trained females during 2 h of cycling at  $65\% \dot{V}O_{2max}$  [26]. In the study by 245 Strauss et al [26], no information was provided on body composition parameters of the 246 female participants. In Şahin et al. [17] (14 days intake, 210 mg anthocyanins per day), 247 enhanced walking-induced fat oxidation was higher with overall body fat % in males. In 248 the present study, we examined whether the enhanced fat oxidation in females was re-249 lated to body mass, body mass index, body fat % in legs, arms and trunk, habitual an-250thocyanin intake, and baseline fat oxidation in rest. The main findings of the present 251 were 1) substantial enhanced walking-induced fat oxidation with intake of New Zealand 252 blackcurrant extract in recreationally active females, and 2) the enhanced walking in-253 duced fat oxidation with intake of New Zealand blackcurrant in recreationally active 254 females was significantly correlated with body mass index and body fat % in legs, arms 255 and trunk. The present study used a dosing strategy of 7-days intake with exercise mo-256 dality and intensity similar to Şahin et al [20]. Şahin et al [20] reported enhanced walk-257ing-induced fat oxidation of with intake of New Zealand blackcurrant extract of 11% in 258 adult males (age:  $26\pm 6$  years, body fat%:  $15\pm 5\%$ ), whereas the present study reports for 259 adult females (body fat%: 31±6%) an increase by 25%. Interestingly, endurance trained 260 males in Cook et al [25] and endurance-trained females in Strauss et al [26] had similar 261 dosing strategies (7-days intake with 210 mg of anthocyanins per day) and provided en-262 hanced 2 hr cycling-induced fat oxidation by 21.5% (males) and 27% (females), respec-263 tively. Therefore, it seems that females are more responsive to New Zealand blackcur-264 rant extract to enhance exercise-induced fat oxidation, and it is also independent of 265 training status. The complexity and the numerous steps involved to alter exercise-in-266 duced fat oxidation and the absence of any biochemical, molecular and structural mark-267 ers in the present study limits the interpretation of the substantial effects on walking-268 induced fat oxidation by intake of New Zealand blackcurrant extract by females. How-269 ever, it is possible that body composition differences between males and females con-270 tributes to the observed effects in the study by Şahin et al [20] and the present study. 271 Future studies should examine the effects of intake of New Zealand blackcurrant extract 272 on exercise-induced fat oxidation in men and women with similar % body fat, albeit it is 273 recognized that the recruitment for such studies will be challenging. Observations that 274 body composition may affect exercise-induced fat oxidation are not consistent. Kerhervé 275 et al [12] observed that body composition differences between women did not affect ex-276 ercise-induced fat oxidation but noted substantial inter-individual differences in normal 277 weight, overweight and obese women. In the present study, the correlation (and signifi-278 cance) was higher for the relationship between body fat % of the legs and the changes in 279 walking-induced fat oxidation. We speculate that there may be heterogenous adipocyte 280 sensitivity, and linked with body fat location, for the effect of anthocyanin-induced me-281 tabolites. If that would be the case, adipocytes in the legs may contribute more to the 282 enhanced exercise-induced fat oxidation than adipocytes in the arms and trunk. This 283 may have consequences for the changes in body composition by long duration intake of 284 anthocyanin-rich blackcurrant supplementation during an exercise intervention. Inter-285 estingly, Isacco et al [13] observed in normal weight pre-menopausal women that exer-286 cise-induced fat oxidation was higher in women with a low abdominal to lower body fat 287 mass ratio. In the present study, we were not able to quantify abdominal fat mass. Fu-288 ture studies should address the effect of long duration intake of anthocyanin-rich black-289 currant supplementation on body composition. 290

Robinson et al [28] reported in recreationally active Caucasian males (n=57, BMI:291 $24.2\pm2.6 \text{ kg}\cdot\text{m}^2$ ) that resting fat oxidation correlated (R=0.55) with exercise-induced maximal fat oxidation. In addition, in the same study,  $\dot{V}O_{2max}$  correlated (R=0.44) with resting293293

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fat oxidation [28]. In the present study, we showed a trend for the correlation between294resting fat oxidation and the change in walking-induced fat oxidation with intake of295New Zealand blackcurrant extract, potentially due that our study only had 12 female296participants. Nevertheless, future work may want to address whether in recreationally297active males and females the  $\dot{V}O_{2max}$ , and thus baseline cardiovascular fitness, may be298predictive of the response to enhanced walking-induced fat oxidation with intake of299New Zealand blackcurrant extract.300

The present study used a convenience sample of recreationally active female partici-301 pants as the primary aim was to examine the effect of intake of New Zealand blackcur-302 rant extract on walking-induced fat oxidation. In a previous study from our group, 303 Matcha green tea drinks in females enhanced walking-induced fat oxidation by 18% [29]. 304 It is possible that the mechanisms for enhanced exercise-induced fat oxidation by differ-305 ent supplementations, i.e. Matcha green tea and New Zealand blackcurrant extract, are 306 not similar. The Matcha green tea study by Willems et al [29] also had recreationally ac-307 tive females as participants and a follow-up study, with the measurement in another 308 laboratory confirmed even higher enhanced fat oxidation of 35% by three weeks Matcha 309 intake [9]. Future studies should examine the combined intake of supplementations of 310 which single use has been shown to enhance exercise-induced fat oxidation. Such infor-311 mation may be useful to inform nutritional strategies for individuals with weight man-312 agement issues. In addition, the effect of supplementation of fat oxidation during rest as 313 well as longer intake duration in females is also important and future work is recom-314 mended. 315

A limitation of the present study was that participants did not record a 48 h food diary 316 for the two visits with taking placebo or New Zealand blackcurrant extract. However, 317 although previous studies [20, 26] had dietary intake recorded, it is unclear how the nu-318 tritional components may have interacted with the intake of blackcurrant anthocyanins 319 and affecting potentially the bioavailability of anthocyanin-derived metabolites. It is the 320 anthocyanin-derived metabolites that are potential linked with adaptative cellular mech-321 anisms. It is concluded that in recreationally active adult females, 7-day intake of NZBC 322 extract substantially enhanced fat oxidation by 30-min of moderate intensity walking 323 exercise. The enhanced walking-induced fat oxidation by intake of NZBC extract in rec-324 reationally active females was body composition-dependent. 325

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Informed Consent Statement: Written informed consent was obtained from all subjects involved in the study. 338

**Data Availability Statement:** The data in this article will be provided by a reasonable request. 341

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## References

- 1. Romijn, J.A.; Coyle, E.F.; Sidossis, L.S.; Rosenblatt, J.; Wolfe, R.R. Substrate metabolism during different exercise intensities in endurance-trained women. *J Appl Physiol* (1985) **2000**, 88(5), 1704-1714. doi: 10.1152/jappl.2000.88.5.1707.
- 2. Lambert, E.V.; Speechly, D.P.; Dennis, S.C.; Noakes, T.D. Enhanced endurance in trained cyclists during moderate intensity exercise following 2 weeks adaptation to a high fat diet. *Eur J Appl Physiol* **1994**, 69(4), 287-293. doi: 10.1007/BF00392032.
- 3. Coggan, A.R.; Kohrt, W.M.; Spina, R.J.; Bier, D.M.; Holloszy, J.O. Endurance training decreases plasma glucose turnover and oxidation during moderate-intensity exercise in men. *J Appl Physiol (1985)* **1990**, 68(3), 990-996. doi: 10.1152/jappl.1990.68.3.990.
- Devries, M.M.; Hamadeh, M.J. Phillips, S.M.; Tarnapolsky, M.A. Menstrual cycle phase and sex influence muscle glycogen utilization and glucose turnover during moderate-intensity endurance exercise. *Am J Physiol Regul Integr Comp Physiol* 2006, 291(4), R1120-R1128. doi: 10.1152/ajpregu.00700.2005.
- 5. Chenevière, X.; Malatesta, D.; Gojanovi, B.; Borrani, F. Differences in whole-body fat oxidation kinetics between cycling and running. *Eur J Appl Physiol* **2010**, 109(6), 1037-1045. doi: 10.1007/s00421-010-1443-5.
- 6. Maunder, E.; Plews, D.J.; Merien, F., Kilding, A.E. Exercise intensity regulates the effect of heat stress on substrate oxidation rates during exercise. *Eur J Sport Sci* **2020**, 20(7), 935-943. doi: 10.1080/17461391.2019.1674928.
- Ruiz-Moreno, C.; Gutiérrez-Hellín, J.; Amaro-Gahete, F.J.; González-García, J.; Giráldez-Costas, V.; Pérez-García, V.; Del Coso, J. Caffeine increases whole-body fat oxidation during 1 h of cycling at Fatmax. J Nutr 2021, 60(4), 2077-2085. doi: 10.1007/s00394-020-02393-z.
- 8. Venables, M.C.; Hulston, C.J.; Cox, H.R.; Jeukendrup, A.E. Green tea extract ingestion, fat oxidation, and glucose tolerance in healthy humans. *Am J Clin Nutr* **2008**, 87(3), 778-784. doi: 10.1093/ajcn/87.3.778.
- 9. Willems, M.E.T.; Fry, H.L.; Belding, M.A.; Kaviani, M. Three Weeks Daily Intake of Match Green Tea Powder Affects Substrate Oxidation during Moderate-Intensity Exercise in Females. *J Diet Suppl* **2021**, 18(5), 566-576. doi: 10.1080/19390211.2020.1811443.
- 10. Cook, M.D.; Myers, S.D. Blacker, S.D. Willems, M.E.T. New Zealand blackcurrant extract improves cycling performance and fat oxidation in cyclists. *Eur J Appl Physiol* **2015**, 115(11), 2357-2365. doi: 10.1007/s00421-015-3215-8.
- Frandsen, J.; Hansen, I.M.D.; Wismann, J.F.; Olsen, M.H.; Brage-Andersen, M.R.; Sahl, R.E.; Hansen, M.; Ingersen, A.; Modvig, J.L.; Schmücker, M.; Grauslund, C.H.; Dela, F.; Larsen, S.; Helge, J.W. Maximal Fat Oxidation Rate is Higher in Fit Women and Unfit Women With Obesity, Compared to Normal-weight Unfit Women. J Clin Endocrinol Metabol 2021, 106(11), e4389-e4399. doi: 10.1210/clinem/dgab473.
- 12. Kerhervé, H.A.; Harvey, L.M.; Eagles, A.N.; McLellan, C.; Lovell, D. Similar rates of fat oxidation during graded submaximal exercise in women of different body composition. *PLoS One* **2020**, 15(11), e0242551. doi: 10.1371/journal.pone.0242551.
- 13. Isacco, L.; Ennequin, G.; Boisseau, N. Effect of Fat Mass Localization on Fat Oxidation During Endurance Exercise in Women. *Front Physiol* **2020** 11, 585137. doi: 10.3389/fphys.2020.585137.
- 14. de Lima, L.P.; de Paula Barbosa, A. A review of the lipolytic effects and the reduction of abdominal fat from bioactive compounds and moro orange extracts. *Heliyon* **2021**, 7(8), e07695. doi: 10.1016/j.heliyon.2021.e07695.
- 15. Nackers, L.M.; Middleton, K.R.; Dubyak, P.J.; Daniels, M.J.; Anton, S.D.; Perri, M.G. Effects of prescribing 1,000 versus 1,500 kilocalories per day in the behavioral treatment of obesity: a randomized trial. *Obesity (Silver Spring)* **2013**, 21(12), 2481-2487. doi: 10.1002/oby.20439.
- 16. Kim, J.; Park, J.; Lim, K. Nutrition Supplements to Stimulate Lipolysis: A Review in Relation to Endurance Exercise Capacity. *J Nutr Sci Vitaminol (Tokyo)* **2016**, 62(3), 141-161. doi: 10.3177/jnsv.62.141.
- 17. Şahin, M.A.; Bilgiç, P.; Montanari, S.; Willems, M.E.T. Daily and Not Every-Other-Day Intake of Anthocyanin-Rich New Zealand Blackcurrant Extract Alters Substrate Oxidation during Moderate-Intensity Walking in Adult Males. *J Diet Suppl* (in press). doi: 10.1080/19390211.2020.1841356.
- 18. Lee, P.H.; Macfarlane, D.J.; Lam T.H.; Stewart, S.M. Validity of the International Physical Activity Questionnaire Short Form (IPAQ-SF): a systematic review. *Int J Behav Nutr Phys Act* **2011**, 8, 115. doi: 10.1186/1479-5868-8-115.
- Bonen, A.; Haynes, F.J.; Watson-Wright, W.; Sopper, M.M.; Pierce, G.N.; Low, M.P.; Graham, T.E. Effects of menstrual cycle on metabolic responses to exercise. *J Appl Physiol Respir Environ Exerc Physiol* 1983, 55(5), 1506-1513.
   392
- Şahin, M.A.; Bilgiç, P.; Montanari, S.; Willems, M.E.T. Intake Duration of Anthocyanin-Rich New Zealand Blackcurrant Extract Affects Metabolic Responses during Moderate Intensity Walking Exercise in Adult Males. J Diet Suppl 2021, 18(4), 406-417. doi: 10.1080/19390211.2020.1783421.
- Neveu, V.; Perez-Jiménez, J.; Vos, F.; Crespy, V.; du Chaffaut, L.; Mennen, L.; Knox, C.; Eisner, R.; Cruz, J.; Wishart, D.; Scalbert,
  A. Phenol-Explorer: an online comprehensive database on polyphenol contents in foods. *Database (Oxford)* 2010, 2010, bap024.
  398
  398
- Willems, M.E.T.; Şahin, M.A.; Cook, M.D. Matcha Green Tea Drinks Enhance Fat Oxidation During Brisk Walking in Females. 399 Int J Sport Nutr Exerc Metab 2018, 28(5), 536-541. doi: 10.1123/ijsnem.2017-0237. 400

23.	Jeukendrup, A.E.; Wallis, G.A. Measurement of substrate oxidation during exercise by means of gas exchange measurements.	401
	Int J Sports Med 2005, 26 Suppl 1, S28-S37. doi: 10.1055/s-2004-830512.	402
24.	Curran-Everett, D.; Benos, D.J. Guidelines for reporting statistics in journals by the American Physiological Society. Adv Physiol	403
	<i>Educ</i> <b>2004</b> , 28(1-4), 85-87. doi: 10.1152/advan.00019.2004.	404
25.	Cook, M.D.; Myers, S.D.; Gault, M.L.; Edwards, V.C.; Willems, M.E.T. Dose effects of New Zealand blackcurrant on substrate	405
	oxidation and physiological responses during prolonged cycling. Eur J Appl Physiol 2017, 117(6), 1207-1216. doi: 10.1007/s00421-	406
	017-3607-z.	407
26.	Strauss, J.A.; Willems, M.E.T.; Shepherd, S.O. New Zealand blackcurrant extract enhances fat oxidation during prolonged cy-	408
	cling in endurance-trained females. Eur J Appl Physiol 2018, 118(6), 1265-1272. doi: 10.1007/s00421-018-3858-3.	409
27.	Elliott-Sale, k.J.; Minahan, C.L.; de Jonge, X.A.K.J.; Ackerman, K.E.; Sipilä, S.; Constantini, N.W.; Lebrun, C.M.; Hackney, A.C.	410
	Methodological Considerations for Studies in Sport and Exercise Science with Women as Participants: A Working Guide for	411
	Standards of Practice for Research on Women. Sports Med 2021, 51(5), 843-861. doi: 10.1007/s40279-021-01435-8.	412

- Robinson, S.L.; Chambers, E.S.; Fletcher, G.; Wallis, G.A. Lipolytic Markers, Insulin and Resting Fat Oxidation are Associated with Maximal Fat Oxidation. *Int J Sports Med* 2016, 37(8), 607-613. doi: 10.1055/s-0042-100291.
- Willems, M.E.T.; Şahin, M.A.; Cook, M.D. Matcha Green Tea Drinks Enhance Fat Oxidation During Brisk Walking in Females. 415 Int J Sport Nutri Exerc Metab 2018, 28(5), 536-541. doi: 10.1123/ijsnem.2017-0237. 416