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Cognitive performance is associated with cerebral oxygenation and peripheral oxygen saturation, but not plasma catecholamines, during graded normobaric hypoxia

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What is the central question of this study?

What are the mechanisms responsible for the decline in cognitive performance following exposure to acute normobaric hypoxia?

What are the main findings and their importance?

We found that 1) performance of a complex central executive task (n-back) was reduced $FiO_2 0.12$; 2) there was a strong correlation between performance of the n-back task and reductions in SpO_2 and cerebral oxygenation; and 3) plasma adrenaline, noradrenaline, cortisol, and copeptin were not correlated with cognitive performance.

Abstract

It is well established that hypoxia impairs cognitive function; however, the physiological mechanisms responsible for these effects have received relatively little attention. This study examined the effects of graded reductions in fraction of inspired oxygen (FiO₂) on oxygen saturation (SpO₂), cerebral oxygenation, cardiorespiratory variables, activity of the sympathoadrenal system (adrenaline, noradrenaline) and hypothalamic-pituitary-adrenal axis (cortisol, copeptin), and cognitive performance. Twelve healthy males (mean [SD], age: 22 [4] yrs, height: 178 [5] cm, mass: 75 [9] kg, FEV₁/FVC ratio: 85 [5] %) completed a 4-task battery of cognitive tests to examine inhibition, selective attention (Eriksen Flanker), executive function (n-back) and simple and choice reaction time (Deary-Liewald). Tests were completed before and following 60 minutes of exposure to FiO₂ 0.2093, 0.17, 0.145, and 0.12.

Following 60 minutes of exposure response accuracy in the n-back task was significantly reduced in FiO₂ 0.12 compared to baseline (82 [9] vs. 93 [5] %; p < 0.001) and compared to all other conditions at the same time point (FiO₂ 0.2093: 92 [3] %, FiO₂ 0.17: 91 [6] %, FiO₂ 0.145: 85 [10] %, FiO₂ 12: 82 [9] %; all p < 0.05). The performance of the other tasks was maintained. Δ accuracy and Δ reaction time of the n-back task was correlated with both Δ SpO₂ (r (9) = 0.66; p < 0.001 and r (9) = - 0.36; p = 0.037 respectively) and Δ cerebral oxygenation (r (7) = 0.55; p < 0.001 and r (7) = - 0.38; p = 0.045 respectively). Plasma adrenaline, noradrenaline, cortisol and copeptin were not significantly elevated in any condition or correlated with any of the tests of cognitive performance. These findings suggest that reductions in peripheral oxygen saturation and cerebral oxygenation, and not increased activity of the sympathoadrenal system and hypothalamic-pituitary-adrenal axis, as previously speculated, are responsible for a decrease in cognitive performance during normobaric hypoxia.

Abbreviations

FiO₂: Fraction of inspired oxygen
SpO₂: Oxygen saturation
SAS: Sympathoadrenal system
HPA axis: Hypothalamic-pituitary-adrenal axis
PFC: Prefrontal cortex
NIRS: Near-infrared spectroscopy
RT: Reaction time
CRT: Choice reaction time
SRT: Simple reaction time
AMS: Acute mountain sickness
ECG: Electrocardiogram

 f_R : Respiratory frequency HHb: Deoxyhaemoglobin O_2 Hb: Oxyhaemoglobin Pa O_2 : Partial pressure of oxygen $P_{ET}CO_2$: Pressure of end tidal carbon dioxide $P_{ET}O_2$: Pressure of end tidal oxygen TSI: Tissue saturation index \dot{V}_E : Minute ventilation V_T : Tidal volume

1. Introduction

The brain requires a constant supply of oxygen in order to support the high rate of ATP production necessary to remain in an electrically active state (Ainslie, Hoiland, & Bailey, 2016; Bailey, Bärtsch, Knauth, & Baumgartner, 2009). Furthermore, as brain activity increases, for example during the performance of a cognitively demanding task, there is an associated rise in the metabolic demands of the neural tissues (Raichle & Gusnard, 2002). As such, a reduction in oxygen availability, as experienced at terrestrial high altitude, can have a detrimental effect on brain function, causing a decline in cognitive performance (Hoiland, Bain, Rieger, Bailey, & Ainslie, 2016).

Despite the growing volume of studies that have examined the effects of hypoxia on cognitive function, the large disparity in methodologies, heterogeneity of cognitive tests, and limited methodological designs (e.g. lack of a control group or normoxic comparison) have resulted in inconsistent findings (Petrassi, Hodkinson, Walters, & Gaydos, 2012; Taylor, Watkins, Marshall, Dascombe, & Foster, 2016; Virués-Ortega, Buela-Casal, Garrido, & Alcázar, 2004; Yan, 2014). Moreover, many of the previous investigations, particularly in aviation research (Petrassi et al., 2012), have focused on establishing an elevation at which cognitive performance decreases, rather than attempting to explore the associated physiological mechanism(s) responsible for a decrease in performance. There is also ambiguity surrounding the type of cognitive tasks that are affected. For example, previous reviews have suggested that decrements in performance were more prominent when completing complex central executive tasks (Petrassi et al., 2012; Taylor et al., 2016; Virués-Ortega et al., 2004; Yan, 2014). Given that more complex central executive tasks activate multiple regions of the brain (Nee, Wager, & Jondies, 2007; Owen, McMillan, Laird, &

Bullmore, 2005), these findings suggest that more demanding cognitive tasks may be more susceptible to hypoxia due to the necessity for greater neural activation. However, using meta-regression analysis, our group has recently shown that the detrimental effects of hypoxia were evident in both complex tasks requiring executive function and also in comparatively simple tasks (e.g. choice and simple reaction time tests) (Terry McMorris, Hale, Barwood, Costello, & Corbett, 2017). That said, our analysis, as with previous reviews, was limited by the number and quality of studies available for inclusion (22 studies comprising only 437 participants). Thus, although it is considered that cognitive performance markedly decreases as the severity of hypoxia increases, the physiological threshold at which impairment begins, the types of cognitive tasks affected and the mechanisms responsible require further investigation.

Recently, Ochi and colleagues demonstrated a decline in the performance of the Stroop task (i.e., a complex central executive task) was associated with decreased SpO₂ during graded normobaric hypoxia (Ochi et al., 2018). These findings indicate arterial oxygen desaturation as a potential physiological factor resulting in hypoxia-induced lowered central executive function. Unfortunately, this investigation did not measure cerebral oxygen delivery (Hoiland et al., 2016). However, cerebral blood flow is unevenly distributed, with greater blood flow directed towards regions of the brain with essential homeostatic roles (e.g. the cerebellum, hypothalamus, thalamus, basal ganglia and brainstem; regions of the brain concerned with cardiorespiratory control) (Binks, Cunningham, Adams, & Banzett, 2007; Willie et al., 2012). Using near infrared spectroscopy (NIRS) the prefrontal cortex (PFC), the primary region of the brain associated with greater activation during the performance of more complex,

central executive tasks (Arnsten & Li, 2005), has been shown to experience a reduction in oxygen saturation during hypoxic exposure (Davranche et al., 2016; Komiyama et al., 2017). Therefore, to elucidate the mechanism(s) responsible for cognitive decline under hypoxic conditions it is also important to investigate the role of cerebral oxygenation in addition to SpO₂.

Another important avenue of exploration that requires further investigation is the relationship between cognitive performance under hypoxia and alterations in catecholamines and stress hormones. Physical and psychological stress, leads to activation of the sympathoadrenal system (SAS) and increased activity of the hypothalamic-pituitary-adrenal (HPA) axis (Schoofs, Preuß, & Wolf, 2008). The initial response of the SAS is mediated via catecholamines, namely adrenaline and noradrenaline. The second, and slightly slower response, consists of activation of the HPA axis and leads to the release of glucocorticoids from the adrenal cortex (e.g. cortisol) (Schoofs et al., 2008). Following acute exposure to hypoxia, resting plasma and / or urinary catecholamines and stress hormones have been shown to significantly increase in comparison to normoxic concentrations (Kamimori et al., 2009; Panjwani, Thakur, Anand, Malhotra, & Banerjee, 2006). As mentioned, the PFC is the primary region of the brain activated during the performance of central executive tasks (Arnsten & Li, 2005). In addition, the performance of simple and choice reaction time type tasks are also associated with activation of the PFC, although they require considerably less neural activation (Barbas, 2000). However, the PFC is highly sensitive to its neurochemical environment and is therefore highly susceptible to stress (Arnsten, 2009). Therefore, these changes have important implications for cognitive performance. Normal function of the PFC is modulated via catecholamine neurotransmitters, in particular dopamine and noradrenaline

signalling in the PFC are critical modulators of prefrontal cognition (Arnsten & Li, 2005). Both dopamine and noradrenaline have an 'inverted U'-shaped influence on PFC function engaging different types of receptors with varying levels of release, such that both inadequate and excessive levels, may cause dysfunction (Arnsten, 2009; Berridge & Spencer, 2016; Terry McMorris, 2016). In addition, it is well established that glucocorticoid hormones released from the adrenal cortex during stress may also impair cognitive performance, with further evidence to suggest that noradrenaline and cortisol may have an additive effect on the decrease in central executive function (Elzinga & Roelofs, 2005). Plasma adrenaline has also been shown to be a predictor of prefrontal cognition, however, this is probably due to the relationship between secretion of adrenaline by the adrenal medulla and the release or noradrenaline in the central nervous system by the hypothalamus via the HPA axis (Terry McMorris, Swain, et al., 2006). Previous research has already examined the influence of alterations in catecholamines on cognitive performance during other forms of stress including sleep deprivation (Terry McMorris et al., 2007), exercise (T. McMorris, Collard, Corbett, Dicks, & Swain, 2008) and heat (Terry McMorris, Swain, et al., 2006), however, to date, there is no empirical evidence directly examining this relationship during hypoxia.

Whilst the aforementioned narrative (Petrassi et al., 2012; Taylor et al., 2016; Virués-Ortega et al., 2004; Yan, 2014) and systematic reviews (Terry McMorris et al., 2017) have provided an insight into this field, the substantial inter-individual variation in the response to hypoxia has made understanding the mechanism(s) by which cognition may be impaired considerably difficult. Therefore, the hypoxia-cognition relationship may be better understood if tested empirically and systematically, whereby participant's cognitive performance and physiological responses are measured

simultaneously during graded levels of hypoxia within a single study. Using a rigorous experimental design, the present study sought to examine simple and complex task performance during exposure to a range of mild-moderate normobaric hypoxic exposures (fraction of expired oxygen [FiO₂] 0.2093, 0.17, 0.145, 0.12) and the concurrent changes in cerebral oxygenation, cardiorespiratory physiology and peripheral biomarkers of SAS and HPA axis activity. We hypothesised that: 1) performance of less complex tasks (i.e. simple and choice reaction time) that require comparatively less neural activation of the PFC and other regions of the brain would be less likely to be reduced following hypoxia than more complex central executive tasks (i.e. Eriksen Flanker test of inhibition and selective attention and n-back) that require comparatively more neural activation of the PFC; and 2) complex task performance would be correlated with changes in SpO₂, cerebral oxygenation and plasma biomarkers associated with SAS and HPA axis activity.

2. Methods

2.1. Ethical approval and Study population

All experimental procedures adhered to the standards set by the latest revision of the declaration of Helsinki, except for registration in a database, and were approved by the Science Faculty Ethics Committee of The University of Portsmouth (project number 2017-025).

A convenience sample of 12 healthy male participants were recruited and participated in this study (mean [SD], age: 22 [4] yrs, height: 178 [5] cm, mass: 75 [9] kg FEV₁/FVC ratio: 85 [5] %. Due to known physiological changes during the menstrual cycle (blood volume, oxygen carrying capacity, ventilation, and body temperature alterations (Takano, 1984)) and the methodological design of the study

(4x conditions) we would not be able to categorically conclude that our findings were attributable to the response of hypoxia or the stage of menses in females; therefore females were not recruited. Following familiarisation with the laboratory and testing procedures, participants provided written, informed consent. Prior to experimentation, participants underwent thorough medical screening to assess for any contraindications to taking part. This included a 12-lead electrocardiogram (ECG), lung function examination and a full blood count. Participants were all nonsmokers and were free of any known cardiovascular, respiratory or neurological disorders. All participants resided at < 500 m and had not spent time altitude for at least three months prior to commencement of the study, including travelling on commercial flight. Participants were instructed to refrain from any strenuous exercise, caffeine and alcohol in the 24 h preceding each visit to the laboratory. In addition, participants were requested to record their dietary intake for 24 h prior to their first visit and to replicate their eating habits for each visit thereafter.

2.2. Study design and Experimental procedures

The study employed a single-blind, randomised, controlled design. Participants were required to visit the laboratory on 6 occasions (health screening, familiarisation and 4 experimental trials). For each experimental trial participants were exposed, using a purpose-built environmental chamber (Sporting edge, Sherfield on Loddon, UK), to normobaric hypoxia for ~60 minutes at one of the following FiO₂ values: 0.2093 (sealevel), 0.17 (equivalent to ~1600 m), 0.145 (~3000 m), and 0.12 (~4500 m). Following extensive pilot work, an exposure time of 60 minutes was used as this time frame was sufficient for all cardiorespiratory variables to plateau. However, we acknowledge that a longer exposure could lead to AMS and mood disturbance and this may lead to a more pronounced decrease in cognitive performance (De Aquino

Lemos et al., 2012). Participants were blinded to the condition they were undertaking and completed an exit questionnaire once all trials were completed to establish whether they were able to correctly identify the conditions for each trial. These questionnaires revealed that the blinding was successful; none of the participants correctly identified the ordering of their conditions. The ambient temperature was maintained at 24 °C and the relative humidity was controlled at 50 % throughout (Komiyama et al., 2015; Sudo et al., 2017). Experimental trials were separated by a minimum of 48 h and conducted at the same time of day. During the exposure, if end tidal O_2 ($P_{ET}O_2$) or end tidal CO_2 ($P_{ET}CO_2$) fell below 45 mmHg and 25 mmHg respectively, for three consecutive breaths, or if SpO₂ went below 65 %, participants were given a supply of normoxic air and subsequently removed from the chamber.

For each trial, the participant was first instrumented outside of the chamber. They then entered the chamber and were connected to a supply or normoxic air (FiO₂ 0.2093). The participant then rested in a semi-reclined position for approximately 10-minutes. The participant then completed the cognitive tests, questionnaires and had a venous blood sample taken (described in 2.3). The supply of normoxic air was then removed and the participant was exposed to the environment (*i.e.* FiO₂ of 0.2093, 0.17, 0.145, or 0.12) within the chamber. Following 60 minutes of exposure, during which the participant remained seated and resting, the cognitive tests and questionnaires were repeated, and another venous blood was sample taken. The participant then left the chamber.

2.3. Measures

2.3.1. Cognitive function

Cognitive function was assessed using a 4-task cognitive battery on a laptop computer (HP Pavilion, China). The battery was designed to examine the performance of both simple and complex cognitive performance. To reduce the occurrence of a learning effect, participants completed each of the tasks 8 times during a familiarisation session, as previously recommended (Taylor et al., 2016). Additional tasks were completed if participants did not meet the following criteria: a) reaction time (RT) variability for the last three trials was ≤ 5 %; b) mean reaction time < 700 ms for n-back and flanker, 300 Ms for simple reaction time (SRT), and 450 for choice reaction time (CRT); and c) response accuracy \geq 90 %. In addition, participants also completed one further practice with each of the tests before the commencement of each experimental trial. Participants were instructed to respond as accurately and as quickly possible. Tasks were presented each time in the order listed below.

2.3.1.1. n-back: Central executive function was assessed using the 3-back version of the n-back number task (Jaeggi, Buschkuehl, Perrig, & Meier, 2010). Participants were presented with a series of letters and required to indicate using the keyboard when the current stimulus was the same as the one presented three trials previously. The task contained 30 trials with each stimulus presented for a maximum of 2000 ms, with a new stimulus presented every 2500 ms. If the participant answered correctly the letter turned green, and if incorrectly it turned red. Accuracy and mean reaction time for correct responses were recorded for analysis.

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2.3.1.2. Eriksen flanker: Inhibition and selective attention (central executive function) were assessed using a modified version of the Eriksen Flanker task (Eriksen, 1995). Participants were presented with 5 letters above a fixation point and required to respond according to the central letter, ignoring the flanker letters. They were instructed to press the 'L' key when the letters 'B' or 'V' appeared and the 'A' key when the letters 'X' or 'C' appeared. The flanker letters could be the same as the target stimuli or representing the pressing of the same button (e.g. an 'X' flanked by 'C' = congruent) or different to the stimulus letter representing pressing of the opposite button (e.g. 'X' flanked by 'B' = incongruent). The interval between the disappearance of the display and the onset of the next was 500 ms. The task included 30 trials in total. Accuracy and mean reaction time for correct responses were recorded for analysis.

2.3.1.3. Simple and choice reaction time

SRT and CRT were measured using the Deary-Liewald Reaction Time Task (Deary, Liewald, & Nissan, 2011). For SRT, a white square set against a blue background was positioned in the centre of the screen. When a black cross appeared within that square the participant had to respond by pressing the space bar as quickly as possible. For CRT, four white squares set against a blue background were positioned in a horizontal line across the middle of the computer screen. When a black cross appeared in one of the squares, the participant was required to press a corresponding key on the keyboard that was in alignment to the position of the squares on the screen. In both tasks, the cross remained on the screen until a button was pressed. The time interval between the response and when the next cross appeared ranged between 100 and 300 ms and was randomised within these

boundaries. Participants were required to complete 20 trials for each of the tasks. Mean reaction time was recorded for analysis, with accuracy also recorded for CRT.

2.4 Physiological measures

2.4.1 Cerebral oxygenation

Cerebral oxygenation was evaluated using the near-infrared spectroscopy (NIRS) technique as previously described (Ando et al., 2013). The NIRS device evaluates the amount of infrared light that effectively traverses an investigated tissue from a transmitter to a receiver. In doing so, the device can detect changes in oxygendependant light absorption by oxy- and deoxy-haemoglobin (Pinti et al., 2018). The NIRS device (Portalite, Artinis Medical, The Netherlands) was used to measure oxygenation of the PFC at wavelengths between 760 and 850 nm. The Portalite was attached to the surface of the left PFC between Fp1 and F3 (international EEG 10-20 system). The measurement site was cleaned with an alcohol wipe and then covered with a transparent film prior to placement of the device. To protect from light interference the device was shielded using a black bandana (Kovalenko, Roskosky, Freedman, & Shuler, 2015). Measurements included an age-dependant differential path-length factor according to manufacturer's guidelines (Duncan et al., 1996). At the beginning of each trial, a resting baseline was established during the initial rest period. All NIRS data were then expressed as changes from baseline (Δ) (Komiyama et al., 2017). The change in oxyhaemoglobin (Δ [O₂Hb]), deoxyhaemoglobin (Δ [HHb]), total haemoglobin (Δ [tHb]), and tissue saturation index (Δ [O2Hb] / Δ [tHb] × 100 %) from baseline were then calculated. In addition, skin blood flow at the forehead was also measured using laser Doppler flowmetry (VMS-LDF2, Moor Instruments, Axminster, UK). These data were used to examine the influence that hypoxia-induced reductions in skin blood flow may have had on the NIRS measurements (Sudo et al., 2017).

2.4.2 Cardiorespiratory variables

Minute ventilation (\dot{V}_E) respiratory frequency (f_R), tidal volume (V_T), and end-tidal pressure of CO₂ ($P_{ET}CO_2$) and O₂ ($P_{ET}O_2$) were measured breath-by-breath using a metabolic cart (Quark CPET, Cosmed, Rome, Italy) as previously described (Neal et al., 2017). Peripheral oxygen saturation (SpO₂) was measured using pulse oximetry on the index finger of the right hand (Nonin 7500, US). Heart rate (HR) was monitored via a 3-lead electrocardiogram (HME, Lifepulse, England). HR and SpO₂ were continually recorded using an analogue to digital data acquisition system (PowerLab 16sp, AD instruments, Castle Hill, Australia).

2.4.3. Blood collection and analysis

Blood was collected via venepuncture from the antecubital fossa at baseline and following 60 minutes of exposure once cognitive tests had been completed. The sample was immediately dispensed into a potassium ethylene diamine tetracetic acid vacutainer and placed on ice. The samples were then centrifuged at 4 °C for 20 minutes at 2500g (Haraeus, Multifuge 3SR Plus, Thermo Electron Scientific instruments, US). The plasma was then aliquoted and stored at -85 °C for later analysis. Commercially available enzyme-linked immunosorbent assays were then used to measure the concentrations of cortisol (Calbiotech, Spring Valley, California, USA), copeptin (Cloud-Clone Corp, Houston, Texas, USA) adrenaline and noradrenaline (both Novusbio, Abingdon, UK). Manufacturer guidelines were followed, and repeated freeze thaw cycles were minimised. Intra-assay coefficient of variation was 5.2 %, 17.0 %, 3.6 % and 4.1 % for cortisol, copeptin, adrenaline and

noradrenaline respectively. Minimum detectable plasma concentrations were 20 ng.mL⁻¹, 12.35 pg.mL⁻¹, 10 pg.mL⁻¹ and 36 pg.mL⁻¹ for cortisol, copeptin, adrenaline and noradrenaline respectively.

2.5. Subjective outcome measures

Acute mountain sickness (AMS) (Lake Louise Score) (Roach et al., 2018), dyspnoea (Modified Borg dyspnoea scale, 0, 'Nothing at all' to 10, 'Shortness of breath so severe you need to stop') (Wilson & Jones, 1989), and mood disturbance (POMS) (Curran, Andrykowski, & Studts, 1995) were self-reported at baseline and following 60 min of exposure.

2.6 Statistical analysis

Physiological data (NIRS and cardiorespiratory measures) were collected for the duration of the trials, however, data were only analysed for the period in which the participants were completing the cognitive tests. The distribution of data was assessed using descriptive methods (skewness, outliers, and distribution plots) and inferential statistics (Shapiro-Wilk test). Main and interaction effects for the physiological data (HR, SpO₂, O₂Hb, HHb, tHb, TSI, \dot{V}_{E} , V_T, *f*_R, P_{ET}CO₂, P_{ET}O₂) and subjective outcome measures (AMS, dyspnoea, mood disturbance) were analysed using a two-way (environment [FiO2: 0.2093, 0.17, 0.145, 0.12] x time [baseline, 60 minutes of exposure]) repeated measures analysis of variance (ANOVA). Cognitive task performance (RT and accuracy) were assessed using the same method, as were adrenaline, noradrenaline, cortisol and copeptin. Where sphericity was violated, degrees of freedom were corrected using the Greenhouse-Geisser method. Where appropriate, post-hoc analysis was performed using the least significant difference (LSD) method. The test of interest for each variable was the significance of the

interaction. Within-participant correlation coefficients were computed for correlations between \dot{V}_E , V_T , f_R , $P_{ET}CO_2$,HR, Δ SpO₂, Δ TSI, Δ cortisol, Δ copeptin, Δ adrenaline, Δ noradrenaline, mood disturbance, dyspnoea, AMS and cognitive task performance (Δ RT and Δ accuracy) using the method described by Bland and Altman [34]. All statistical analyses were performed using SPSS (Statistical Package for the Social Sciences), version 24.0 (SPSS Inc, Chicago, IL, USA). Data are presented as mean [SD] unless otherwise stated. Statistical significance was accepted at p < 0.05.

3. Results

One participant was removed from the chamber in $FiO_2 \ 0.12 \ (P_{ET}O_2 \ fell \ below \ 45 \ mmHg)$. Therefore, the following analyses are for the 11 participants that successfully completed all 4 experimental trials.

3.1. Cognitive function

Performance for each of the cognitive tasks are displayed in Table 1. Individual responses to the tasks (Δ RT and Δ accuracy) are shown in Figure 1. Analysis revealed no significant changes in RT for any of the tasks (all *p* > 0.05). Accuracy was maintained in each of the tasks with the exception of the n-back task, where there was a significant interaction effect ($F_{(3,30)} = 9.157$; *p* < 0.001). Following 60 minutes of exposure accuracy was significantly lower in FiO₂ 0.12 compared to baseline (*p* < 0.001) and was significantly reduced compared to FiO₂ 0.2093 (*p* = 0.004) and FiO₂ 0.17 (*p* = 0.002) at the same time point. Accuracy was also significantly reduced in FiO₂ 0.145 following 60 minutes of exposure compared to FiO₂ 0.2093 (*p* = 0.049), however, it was not significantly different to baseline (*p* > 0.05).

3.2. Physiological measures

Participants commenced each of the experimental trials from a resting baseline, with no significant differences between trials (all p > 0.05, Table 2).

3.2.1. Cerebral oxygenation

Due to equipment malfunction the following NIRS analyses are for 9 participants only (Figure 2B-E). There was a significant interaction effect for ΔO_2 Hb (F_(3,24) = 7.304; *p* = 0.001), Δ HHb (F_(3,24) = 43.360; *p* < 0.001) and Δ TSI (F_(3,24) = 12.810; *p* < 0.001). In each condition, with the exception of FiO₂ 0.2093, there was a significant reduction in ΔO_2 Hb (all *p* < 0.05) and a reciprocal increase in Δ HHb compared to baseline (all *p* < 0.05). Following 60 minutes of exposure, ΔO_2 Hb was significantly lower and Δ HHb was significantly higher with each reduction in FiO₂ (all *p* < 0.05). Similarly, Δ TSI was significantly reduced following 60 minutes of exposure compared to baseline in all conditions except for FiO₂ 0.2093 (all *p* < 0.05). In FiO₂ 0.12, Δ TSI was significantly lower than each of the other conditions following 60 minutes of exposure (all *p* < 0.05). There was no significant interaction effect for Δ tHb (*p* = 0.055). The correlation between Δ skin blood flow and ΔO_2 Hb was not significant (*r* (7) = -0.23, *p* = 0.237).

3.2.2. Cardiorespiratory response

There were no significant changes in \dot{V}_E , V_T or f_R across conditions (all p > 0.05; Table 2). Compared to baseline, both $P_{ET}CO_2$ and $P_{ET}O_2$ were significantly reduced following 60 minutes of exposure in all conditions with the exception of FiO₂ 0.2093 (all p < 0.05). As expected, there was a significant reduction in $P_{ET}O_2$ with each reduction FiO₂ (FiO₂ 0.2093 > 0.17 > 0.145 > 0.12; all p < 0.05). A significant interaction effect for SpO₂ was observed ($F_{(3,30)} = 107.902$; p < 0.001; Figure 2A). There was a significant reduction in SpO₂ in all conditions following 60 minutes of exposure compared to baseline, with the exception of FiO₂ 0.2093 (all p < 0.05). As designed, there was a concomitant decrease in SpO₂ with each reduction in FiO₂ (0.2093 > 0.17 > 0.145 > 0.12; all p < 0.05). A significant interaction was detected for HR ($F_{(3,30)} = 8.640$; p < 0.001). Compared to baseline, there was a significant reduction in HR in FiO₂ 0.2093 (p < 0.001) and a significant increase in FiO₂ 0.12 (p = 0.023). Following 60 minutes of exposure, HR was significantly higher in both FiO₂ 0.145 and 0.12 compared to 0.17 and 0.2093 (p < 0.05).

3.3. Biomarkers

There were no between environment differences observed for any of the biomarkers following 60 minutes of exposure (all p < 0.05). There were also no significant changes from baseline for any of these measures (Figure 3A-D).

3.4. Subjective outcome measures

The questionnaire data indicated no significant change in mood or dyspnoea. In addition, whilst some of the participants reported various symptoms of AMS (headache, feeling tired), none were classified as having AMS according to the Lake Louise AMS diagnosis criteria (Table 2).

3.5. Correlational analysis

Correlational analysis revealed a strong, positive correlation between Δ accuracy in the n-back task and both Δ SpO₂ (r (9) = 0.66; *p* < 0.001; figure 3B) and Δ TSI (r (7) = 0.55; *p* < 0.001; Figure 4D). We also observed a moderate negative correlation between Δ reaction time for the n-back task and both Δ SpO₂ (r (9) = -0.36; *p* = 0.037; Figure 4A) and Δ TSI (r (7) = - 0.38; *p* = 0.045; figure 4C). There was also a very strong, positive correlation between Δ TSI and Δ SpO₂ (r (7) = 0.77; *p* < 0.001). No other significant correlations were observed.

4. Discussion

This study is the first to systematically examine the relationship between cognitive performance and cerebral oxygenation, peripheral oxygen saturation. sympathoadrenal system and hypothalamic-pituitary-adrenal axis activity during exposure to graded mild- to moderate- levels of normobaric hypoxia. The principal novel findings from this study are: 1) accuracy on the n-back task (indicative of central executive function) was reduced at FiO₂ 0.12; 2) central executive function, assessed using accuracy and reaction time on the n-back task, was significantly correlated with reductions in SpO₂ and cerebral oxygenation; and 3) peripheral levels of adrenaline, noradrenaline, cortisol and copeptin were not significantly elevated and were *not* correlated with the cognitive task performance assessed. Collectively, these findings suggest that central executive performance on complex cognitive tasks (n-back) decreases during rest at a simulated altitude equivalent to ~4500 m. This decrease in performance is associated with reductions in peripheral and cerebral oxygenation, but not sympathoadrenal system or hypothalamic-pituitaryadrenal axis activity as previously speculated in the literature.

In partial agreement with our first hypothesis, we observed maintained performance in SRT and CRT and a decline in performance of the n-back task (Table 1; Figure 1). However, in contrast to our hypothesis, we found no decrements in the performance of the flanker task. Firstly, it is unsurprising that we only observed a significant decrement in performance in the FiO_2 0.12 condition. Whilst several studies have reported a decrease in cognitive performance at altitudes (or simulated altitudes) between 2000-3500 m (Terry McMorris et al., 2017; Petrassi et al., 2012), decrements in performance are more pronounced once elevation exceeds 4000 m. Thus, despite a significant reduction in cerebral oxygenation in each of the hypoxic conditions, it was not until desaturation reached ~5% that cognitive performance was significantly reduced.

Despite our recent meta-analysis (McMorris et al., 2017) suggesting that both simple and complex task performance are reduced following hypoxic exposure, we felt that due to the limited availability and variable quality of evidence on the topic, examination of both simple and complex task performance was required. Our hypothesis that simple task performance (SRT and CRT) would be less likely to be reduced following hypoxia than complex task performance (n-back and Eriksen flanker) was therefore based on the reported dysfunction of the PFC due to alterations in its neurochemical environment following psychophysiological stress (Arnsten, 2009; Arnsten & Li, 2005; Berridge & Spencer, 2016), and the requirement of several other brain regions that may also be susceptible to hypoxia. An interesting finding to emerge from the current study was that although a decrease in n-back performance was observed, performance of the flanker task was maintained. Whilst both the n-back and flanker tasks are considered to be complex tasks that place a load on central executive function, the tasks are considerably different, and as such, may also therefore require activation of different regions of the brain. A metaanalysis of functional neuroimaging studies focusing on non-verbal n-back tasks (Owen, McMillan, Laird, & Bullmore, 2005) similar to that used in the current study, revealed brain activation in six key frontal and parietal regions, namely the right dorsolateral prefrontal, lateral premotor and posterior parietal cortex, a set of regions

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that have been described as the spatial attention network. However, a meta-analysis of flanker studies observed brain activation in the right dorsolateral prefrontal cortex with additional activation in the right insula (Nee, Wager, & Jondies, 2007). Thus, in comparison to the flanker, the n-back appears to require activation of several different brain regions of which may be more susceptible to increased levels of hypoxia, however this proposal requires further investigation.

Although we did not measure the central executive (attention) demand of either task, it may be that the demands for the 3-back test are greater than those for the flanker. Scharinger et al. (Scharinger, Soutschek, & Schubert, 2015), using a combined n-back/flanker task, examined the effect of increased demand on the n-back factor of the task on performance of the flanker factor. They found an underadditive interference effect, i.e. facilitation of performance on the flanker factor when the n-back factor increased from 0- and 1-back to 2-back. This was the case for behavioural, electroencephalograph and pupillometry data. The authors interpreted this as showing that the activation of networks common to both tasks was so great during the 2-back test that the flanker task required no extra resources unlike in the 0-back and 1-back conditions. This does suggest that 3-back may require a greater attention demand than the flanker task.

In a study that investigated cerebral oxygenation during hypoxia, Subudhi et al. (2009) reported that the extent of deoxygenation was greatest in the prefrontal areas, however, patterns of deoxygenation were similar across multiple regions of the brain (i.e. prefrontal, premotor, and motor cortices (Subudhi, Miramon, Granger, & Roach, 2009). Thus, although we only measured oxygenation of the PFC, it is reasonable to suggest that hypoxia resulted in a similar reduction in oxygenation across the multiple brain regions activated during the n-back task. Therefore, we

speculate that the reduction in n-back performance was caused by (i) reduced oxygenation of the PFC and the other regions of the brain associated with n-back performance, and (ii) also, or in isolation, the addition of the hypoxic stressor altered the interaction between these regions. This may potentially explain the lack of relationship between the less complex tasks and both SpO₂ and cerebral oxygenation. Future research is warranted to explore these proposed physiological mechanism(s) as the current research suggests that the effects of hypoxia on cognitive performance is related to the degree of neural activation required to complete the task.

In partial agreement with our second hypothesis, n-back performance (RT and accuracy) was significantly correlated with both Δ SpO₂ and Δ TSI (Figure 3). These findings support those of Ochi et al., (2018) who reported that SpO₂ was associated with performance of the Stroop task in hypoxic conditions similar to those used in this study (FiO₂ = 0.165, 0.135, 0.105; r = -0.293, p < 0.01). Our data further extends the findings of Ochi et al., (2018) by providing the first experimental evidence, using a rigorous, within subject design, that n-back performance (a task requiring executive function) is associated with both peripheral arterial oxygen desaturation and oxygenation of the PFC. Oxygenation of the PFC is also affected by cerebral vasoconstriction caused by hypoxia-induced hyperventilation (Ogoh et al., 2014; Steinback & Poulin, 2008). Although we observed a significant alteration in P_{ET}CO₂, the magnitude of change was small (3 mmHg). Furthermore, there was also no significant correlation between P_{ET}CO₂ and TSI. In contrast, we observed a very strong positive correlation between SpO₂ and TSI (r (7) = 0.77; p < 0.001), suggesting that reduced arterial oxygenation had a greater role in deoxygenation of the PFC than hyperventilation-induced restriction of cerebral blood flow. Furthermore, it is also important to note that we observed no relationship between Δ skin blood flow and Δ O₂Hb (*p* = 0.237) suggesting the NIRS device was providing a measure that accurately reflected changes in cerebral oxygenation and not skin blood flow.

Despite observing a considerable reduction in both SpO₂ and cerebral oxygenation we found no significant alterations in sympathoadrenal system or hypothalamicpituitary-adrenal axis activity. Adrenaline, noradrenaline, cortisol, and copeptin values were within the expected range for normal resting values following 60 minutes of exposure and no relationship between any of the biomarkers and cognitive performance was observed. These findings support those of an earlier review that found only trivial changes in plasma catecholamines following hypoxic exposure (Rostrup, 1998). The minimal increases in cortisol, the most widely accepted measure of HPA axis activity (Costello et al., 2018; Morgan et al., 2000), \dot{V}_{E} , and heart rate suggest that the participants experienced only minor levels of physiological strain even in the most severe hypoxic exposure (Table 2). It is of great interest therefore that we still observed a significant decrease in cognitive performance on the n-back task. Although our findings suggest there was not an increase in noradrenaline, it is possible that as catecholamines do not cross the blood brain barrier, that this was not reflective of what was occurring in the brain (Terry McMorris, Harris, et al., 2006). However, we included the measurement of plasma concentrations as circulating adrenaline and noradrenaline activate βadrenoceptors on the vagus nerve, which feeds back to the nucleus of the solitary tract and activates catecholaminergic neurons in the medulla and pons (Terry McMorris, 2016). These initiate the synthesis and release of noradrenaline from the locus coeruleus which projects to the PFC and indeed throughout the brain (Terry

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McMorris, 2016). In addition, the turnover of catecholamines is altered in response to hypoxia due to the requirement of oxygen during synthesis, release and metabolism (Gibson, Pulsinelli, Blass, & Duffy, 1981). It is therefore plausible that although plasma noradrenaline concentration was unchanged, its turnover may have been. This may be indicative of a similar response in the brain. Furthermore, cortisol does cross the blood brain barrier (Levine, Zagoory-Sharon, Feldman, Lewis, & Weller, 2007) and so peripheral concentrations may be indicative of central levels. That cortisol also showed no significant differences between conditions supports the claim that hypoxia disrupts neurochemical turnover in the brain (Gibson et al., 1981). Nevertheless, future studies that measure changes in plasma concentrations of metabolites noradrenaline of and dopamine such 3,4as methoxyhydroxyphenylglycol and homovanillic acid, which do cross the blood brain barrier may provide greater understanding of the mechanistic processes underpinning cognitive performance.

Further to the future directions considered thus far in the discussion, the experimental design used in the present study and associated findings have created interesting hypotheses that warrant further investigation. Firstly, whilst this study has expanded on the work of Ochi et al. (2018) by including a measure of cerebral oxygenation and plasma biomarkers associated with SAS and HPA axis activity, we did not measure cerebral blood flow. This is an important variable which should be considered in future investigations (Ogoh et al., 2014). A potential limitation of this study may have been that participants only rested for 10 minutes prior to the start of the experiment. Consequently, ventilation may have been elevated at the beginning of the experiment and therefore explain why we observed no changes in ventilation after 60 minutes of exposure. This appears to be further confirmed by the decrease

in heart rate we observed in the FiO₂ 0.2093 condition. Future studies may want to ensure a greater rest time is given prior to commencement. In order to exclude any confounding factors brought about by exercise, the current study was conducted whilst participants were at rest, however, it is well accepted that cognition may be improved during moderate intensity exercise (Chang, Labban, Gapin, & Etnier, 2012; Terry McMorris, 2016). Furthermore, there is emerging evidence to suggest that the beneficial effects of exercise may outweigh the negative effects of hypoxia, dependent upon the severity of hypoxia and the intensity of the exercise (Komiyama et al., 2017). With a high proportion of activities taking place at altitude involving exercise (e.g. skiing, military operations, mountain rescue) future studies, using a similar robust methodological construct, should examine changes in cognitive performance when exercising. Also, hypoxia is seldom experienced in isolation (M. Tipton, 2012; M. J. Tipton, 2016) and is often encountered alongside various other stressors such as fatigue, dehydration, sleep deprivation, or other environmental conditions such as the cold or the heat. As this is one of the first studies attempting to delineate the physiological mechanism(s) responsible for cognitive decline in hypoxia, further research is required to examine the impact of combining these additional stressors with hypoxia. It is also important to note that the rigorous health screening undertaken by participants in this study meant that all participants were in excellent health and as such, this may not have been a true representation of the average population. This may also explain why previous studies with no such prescreening have experienced greater decrements in performance at lower levels of hypoxia. Finally, considering the influence of sex on cerebral function (Robison, Gannon, Salinero, & Zuloaga, 2019), it is necessary for future studies to confirm these findings in females.

In conclusion, hypoxia-induced decreases in central executive performance are associated with reductions in both SpO₂ and cerebral oxygenation, but not plasma adrenaline, noradrenaline, cortisol, or copeptin. Nevertheless, the substantial majority of the inter-individual variance in cognitive performance following exposure to hypoxia remains unaccounted for and future studies should seek to increase understanding of the physiological factors contributing to a decrease in central executive performance.

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-5-

-10

Baseline

60 min exposure



TIC E

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Table 1. Reaction time and accuracy for the cognitive tests at baseline and following60 minutes of exposure (n=11).

		FiO ₂							<i>p v</i> alue		
	0.2093		0.17		0.145		0.12		FiO	Ti me	Interac tion
	Basel ine	Expos ure	Basel ine	Expos ure	Basel ine	Expos ure	Basel ine	Expos ure			
Simple reactio n time Reactio	257	253	244	250	261	269	241	282	0.0	0.0	0.440
n time (ms) Choice reactio n time	[26]	[14]	[13]	[19]	[28]	[11]	[15]	[28]	28	24	0.142
Reactio n time (ms)	381 [48]	370 [50]	353 [33]	356 [36]	352 [37]	370 [50]	358 [43]	380 [52]	0.1 53	0.7 00	0.159
Accurac y (%) Eriksen Flanker Congru	98 [5]	95 [7]	96 [6]	97 [3]	95 [4]	97 [3]	97 [3]	96 [3]	0.9 40	0.8 37	0.172
ent reaction time (ms) Congru	532 [91]	517 [88]	527 [75]	524 [76]	523 [77]	510 [87]	515 [78]	517 [59]	0.7 65	0.4 06	0.831
ent accurac y (%) Incongr	99 [2]	98 [5]	96 [9]	96 [6]	96 [7]	96 [4]	99 [3]	95 [6]	0.2 53	0.2 56	0.622
uent reaction time (ms) Incongr	544 [74]	550 [68]	568 [83]	558 [101]	552 [85]	543 [84]	534 [61]	556 [64]	0.5 04	0.7 95	0.184
uent accurac y (%)	94 [9]	92 [12]	96 [5]	93 [6]	93 [11]	93 [9]	94 [7]	92 [6]	0.8 75	0.3 30	0.671
N-back											
Reactio n time (ms)	598 [125]	578 [95]	614 [146]	631 [106]	616 [109]	636 [109]	613 [129]	680 [131]	0.5 45	0.1 64	0.204
Accurac y (%)	90 [6]	92 [3]	89 [8]	91 [6]	86 [9]	85 [10] ^ь	93 [5] ^d	82 [9] ^{abc}	0.1 67	0.1 52	< 0.001

Values are mean [SD]. ^a significantly different (p < 0.05) to the baseline value in the same environment. ^b significantly different (p < 0.05) to same time point in FiO₂ 0.2093. ^c significantly different (p < 0.05) to same time point in FiO₂ 0.17. ^d significantly different (p < 0.05) to same time point in FiO₂ 0.145.

Table 2. Cardiorespiratory responses and subjective outcome measured during the cognitive tasks at baseline and after 60 minutes of exposure (n = 11).

$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$					Fi	O ₂				p value			
$ \begin{array}{ c c c c c c c c c c c c c c c c c c c$		0.2	2093	0	.17	0.	145	0	.12	FiO 2	Ti me	Interac tion	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $		Basel ine	Expos ure	Basel ine	Expos ure	Basel ine	Expos ure	Basel ine	Expos ure				
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	HR (b.min ⁻¹)	70 [9]	65 [9]ª	69 [12]	69 [12]	72 [10]	74 [10] ^{bc}	71 [11]	76 [13] ^{abc}	0 .04 9	0.6 17	< 0.001	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	V _E (L.min⁻¹)	14.2 [1.2]	12.3 [2.0]	13.5 [2.4]	12.5 [2.6]	14.1 [1.8]	13.8 [2.6]	13.4 [2.9]	13.4 [3.6]	0.6 93	0.0 38	0.121	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	V⊤ (L BTPS)	0.801 [0.139]	0.892 [0.423]	0.754 [0.132]	0.807 [0.276]	0.818 [0.173]	0.836 [0.122]	0.809 [0.156]	0.913 [0.247]	0.5 40	0.1 75	0.262	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	^f _R (breaths. min ⁻¹)	18.0 [2.3]	15.5 [4.3]	18.1 [2.4]	16.4 [3.8]	17.5 [2.6]	16.8 [4.2]	16.9 [3.6]	15.2 [4.6]	0.2 02	0.0 16	0.583	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	P _{ET} CO ₂ (mmHg)	38 [2]	37 [2]	38 [2]	37 [2]ª	38 [2]	37 [2]ª	38 [2]	35 [4] ^{abc}	0.1 76	0.0 01	0.007	
AMS 0 [1] 1 [2] 0 [1] 1 [2] 0 [0] 0 [2] 1 [1] 2 [2] $\begin{array}{c} 0.2 \\ 67 \\ 04 \\ 0.09 \\ 0.0 \\ 0.0 \\ 0.0 \\ 0.09 \\ 0.099 \\ 0.00 \\ 0.186 \\ 0.00 \\ 0.0 \\ 0.0 \\ 0.0 \\ 0.0 \\ 0.099 \\ 0.00 \\ 0.0 \\ 0.099 \\ 0.00 \\ 0.0 \\ 0.099 \\ 0.00 \\ 0.0 \\ 0.099 \\ 0.00 \\ 0.0 \\ 0.09 \\ 0.099 \\ 0.00 \\ 0.00 \\ 0.0 \\ 0.00 \\ 0.0 \\ 0.09 \\ 0.099 \\ 0.00 \\ 0.0 \\ 0.00 \\ 0.0 \\ 0.09 \\ 0.09 \\ 0.09 \\ 0.00 \\ 0.0 \\ 0.00 \\ 0.0 \\ 0.00 \\ 0$	P _{ET} O₂ (mmHg)	109 [2]	108 [2]	104 [5]	78 [3] ^{ab}	103 [6]	61 [4] ^{abc}	101 [7]	48 [2] ^{abcd}	< 0.0 01	< 0.0 01	< 0.001	
Dyspnoe 0 [0] 0 [1] 0 [0] 0 [1] 0 [0] 0 [1] 0 [0] 0 [1] 0 [0] 1 [2] 0 [0] 1 [2] 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0 0	AMS	0 [1]	1 [2]	0 [1]	1 [2]	0 [0]	0 [2]	1 [1]	2 [2]	0.2 67	0.0 04	0.694	
^{Mood} Disturba 34 [6] 39 [11] $\stackrel{33}{[13]}$ 44 [23] $\stackrel{36}{[10]}$ 43 [24] $\stackrel{37}{[13]}$ 42 [26] $\stackrel{0.1}{18}$ $\stackrel{0.0}{02}$ 0.186 Cardiorespiratory values are are mean [SD]. HR, heart rate; \dot{V}_{E} , minute ventilation; f_{R} , respiratory frequency; V _T , tidal volume; P _{ET} CO ₂ , end-tidal partial pressure of carbon dioxide; P _{ET} O ₂ , end-tidal partial pressure of carbon dioxide; P _{ET} O ₂ , end-tidal partial pressure of carbon dioxide; Net endities and the same environment. ^b significantly different ($p < 0.05$) to the baseline value in the same environment. ^b significantly different ($p < 0.05$) to same time point in FiO ₂ 0.2093. ^c significantly different ($p < 0.05$) to same time point in FiO ₂ 0.145.	Dyspnoe a	0 [0]	0 [1]	0 [0]	0 [1]	0 [0]	0 [1]	0 [0]	1 [2]	0.0 41	0.0 78	0.099	
Cardiorespiratory values are mean [SD]. HR, heart rate; \dot{V}_{E} , minute ventilation; f_{R} , respiratory frequency; V_{T} , tidal volume; $P_{ET}CO_2$, end-tidal partial pressure of carbon dioxide; $P_{ET}O_2$, end-tidal partial pressure of carbon dioxide. Subjective outcome measures are median [IQR]. AMS; acute mountain sickness. ^a significantly different ($p < 0.05$) to the baseline value in the same environment. ^b significantly different ($p < 0.05$) to same time point in FiO ₂ 0.2093. ^c significantly different ($p < 0.05$) to same time point in FiO ₂ 0.17. ^d significantly different ($p < 0.05$) to same time point in FiO ₂ 0.05) to same time point in FiO ₂ 0.145.	Mood Disturba nce	34 [6]	39 [11]	33 [13]	44 [23]	36 [10]	43 [24]	37 [13]	42 [26]	0.1 18	0.0 02	0.186	
	Cardiore requent partial p nountai significa same tin	espirator cy; V _T , f ressure n sickne ntly diffe ne point	ry values tidal volui e of carbo ess. ^a sign erent ($p <$: in FiO ₂ 0	are are me; P _{ET} C on dioxide ificantly o 0.05) to .17. ^d sign	mean [SE :O ₂ , end-t e. Subjec different (µ same time nificantly c	D]. HR, h idal parti tive outc o < 0.05) e point in different (eart rate; ial pressu ome mea to the bas $FiO_2 0.20$ p < 0.05)	V _E , min ure of car asures ar seline val 093. ^c sigr to same t	ute ventila bon dioxi e median ue in the nificantly o time point	ation; <i>f</i> ide; P _{E⁻ i [IQR]. same e different in FiO₂}	R, res O ₂ , e AMS nviror (<i>p</i> < 0.14	spiratory end-tidal 5; acute nment. ^b 0.05) to 5.	